

GIANT
Gene therapy: an Integrated Approach for Neoplastic Treatment

LMU

„ Mechanisms of non-viral gene delivery “

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nsm 14 06 2008 **CeNS**
nanosystems initiative munich Center for NanoScience Ludwig-Maximilians-Universität

nsm Programmed Nanosystems – ‘Synthetic Viruses’ **LMU**

- Specific delivery and effective drug release
- Timely operation of multiple tasks

‘Smart’ Nanosystems **Extra & Intra - Cellular Pathways**

Shielding function
(blood circulation)
Targeting function
Triggered release
(at target site/cell)

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EGF receptor targeted delivery

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nsm Interactions with Cells **LMU**

DNA/PEI **DNA/PEI/PEG/EGF**

actin filaments **EGF receptor**

HSPG **early endosome** **early endosome**

DNA/PEI/PEG **late endosome**

nucleus

motor protein
microtubule

nsm Observation of Single Polyplexes on Living Cells **LMU**

Cell with actin-GFP labeled cytoskeleton Cy5-labeled polyplexes

Christoph Bräuchle
Karla de Bruin
Nadia Ruthardt

10 μm

nsm EGF Polyplex Tracking: Three Phases **LMU**

3 μm

1 μm

the color codes for corresponding time points in the trajectory

instantaneous velocity [μm/s]

time [s]

three phases observed:

phase I: slow directed motion

phase II: diffusion

phase III: directed motion along microtubules

tracked for 4:30 min at a frame rate of 300 ms

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0 sec

Phase I
PEI particles on actin-GFP cell

recorded at 5 second intervals for 54 min, 500 ms acquisition time, 75x time lapse

Phase II
(occasionally actin comets)

60 x time lapse

HUH-7 cells / GFP-tagged actin

K. deBruin et al, *Mol. Ther.* 2007

nsm Particle Internalization: Quenching Assay **LMU**

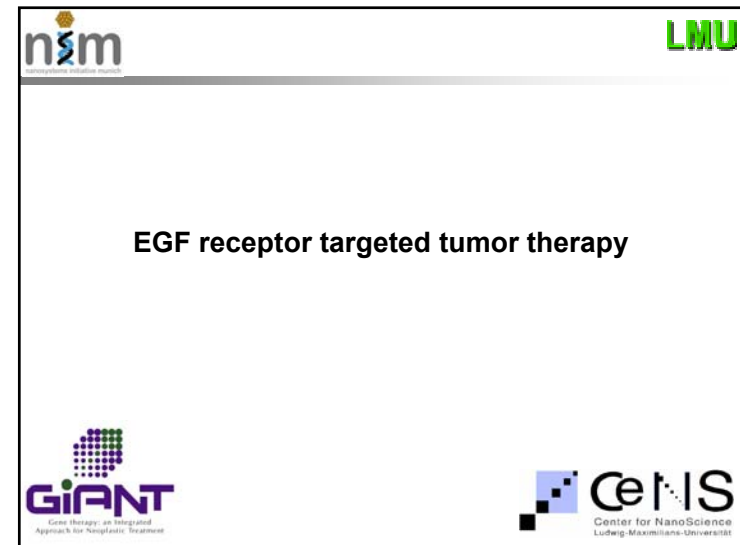
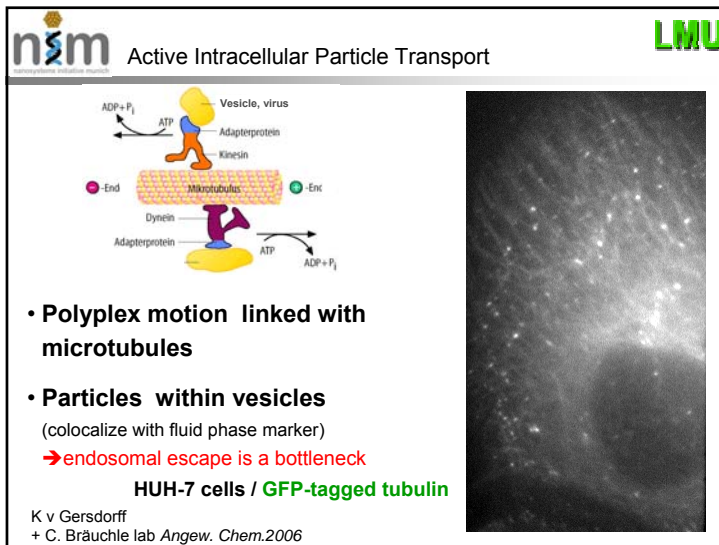
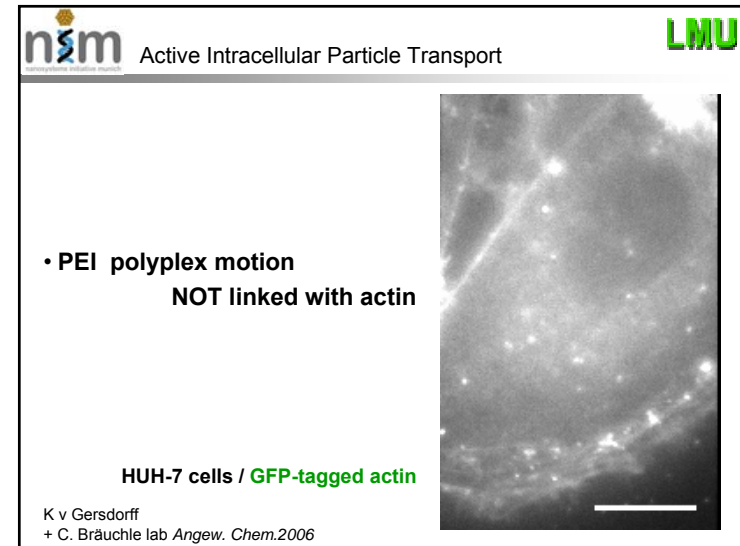
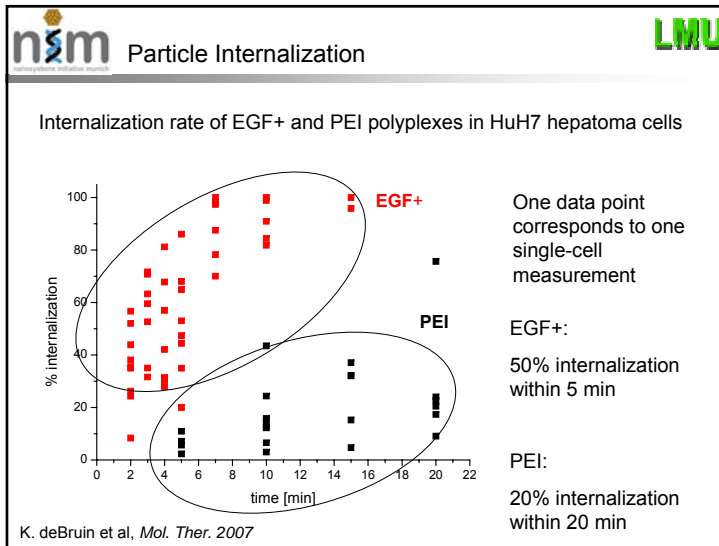
10 μm

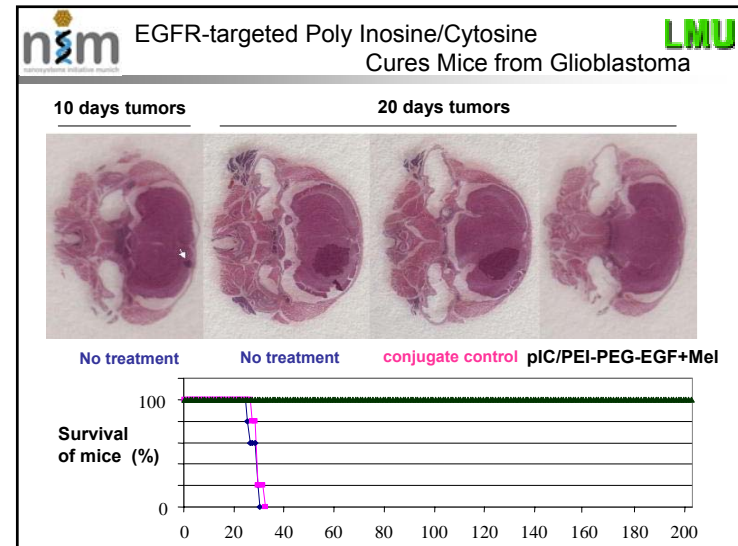
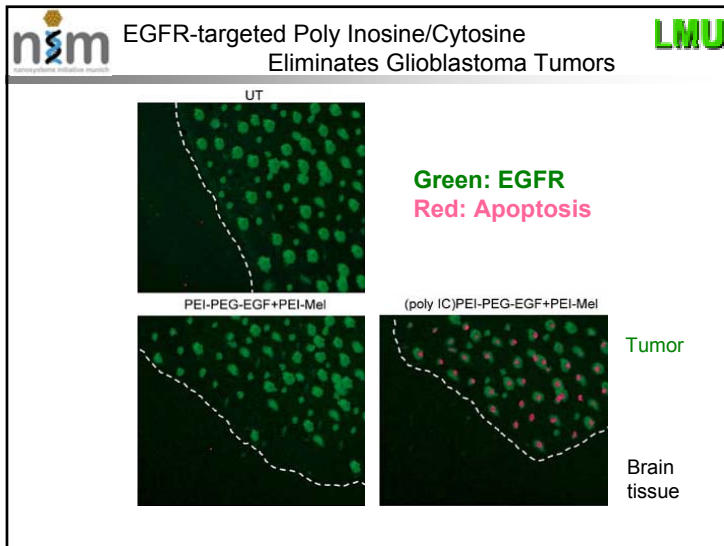
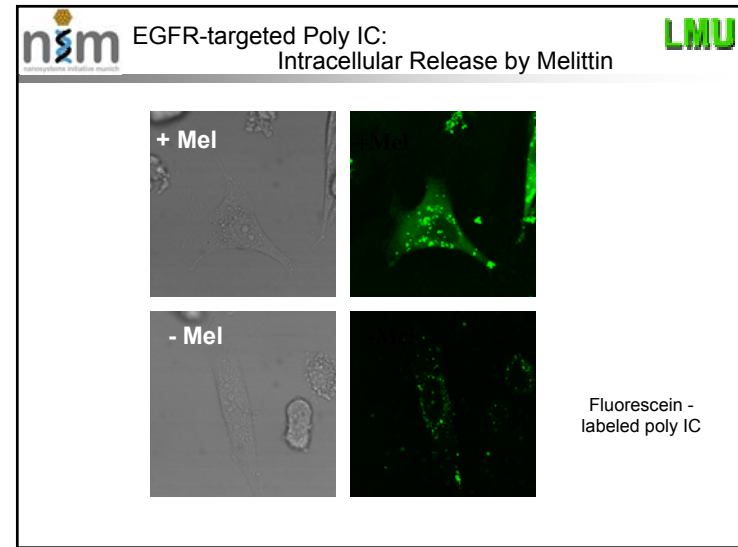
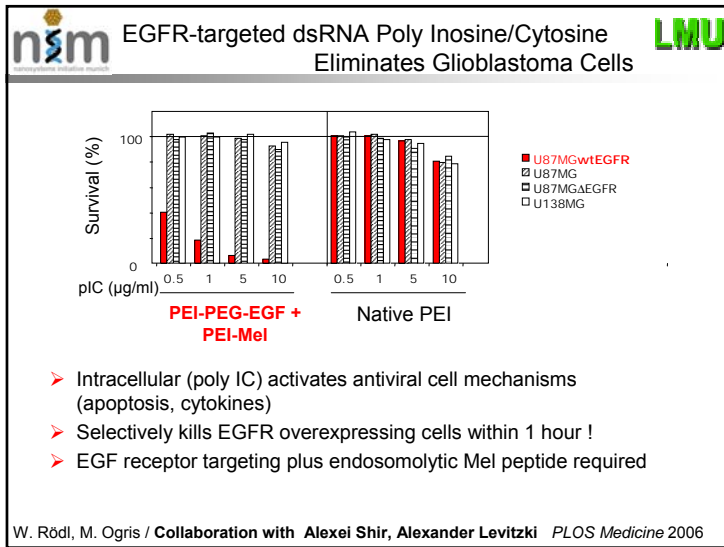
fluorescence intensity [a.u.]

time [s]

— particle 1
— particle 2
— particle 3
— particle 4
— particle 5
— particle 6
— threshold

Quenching of Cy3 fluorescence with the membrane-impermeable dye trypan blue
→ only extracellular particles will be quenched





EGFR-targeted poly (I•C):
Other EGFR - overexpressing Tumors

► “Tetra-conjugate” MPPE more effective than mixture of two conjugates

► Complete elimination of established breast cancer and adenocarcinoma xenografts

The top graph shows survival percentage for MDA-MB, A431, and U87MG cell lines under different treatment conditions (UT, PEI₂₅-PEG-EGF+PEI₂₅-Mel, and Mel-PEI₂₅-PEG-EGF) at 1, 5, and 10 µg/ml. The bottom graph shows tumor size (mm³) over time for A431 and MDA-MB-468 xenografts, comparing UT, MPPE, and poly ICM/PPE treatments.

W. Rödl, M. Ogris / A. Shir, A. Levitzki *PLOS Medicine* 2006

Improved Polyplexes:

- Better Defined Polymers
- Better Endosomal Escape
- Extracellular stabilization
- Intracellular disassembly

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Biodegradable (Ester, Disulfide, Amide-based) Polyamines

The scheme shows the synthesis of polyamines from oligoamines using DTBP and DSP. It also illustrates the formation of HD linkers via HD at 20°C and HD linkers via 60° ester aminolysis. The resulting structures include IP and SP linkers.

Triethylenetetramine	TT (N=4)
Tetraethylenepentamine	TP (N=5)
Pentaethylenhexamine	PH (N=6)
Oligoethylenimine 800	OEI (N~18)

$H_2N-(CH_2)_3-NH-(CH_2)_4-[NH-(CH_2)_3]_x-NH_2$

Spermidine	SD (N=3)
Spermine	SP (N=4)

Julia Klöckner et al *Eur. J. Pharm. Sci* 2006
Peter Tarcha et al *Biomaterials* 2007

Pseudodendritic Polyamines: Better Defined

The scheme shows the synthesis of pseudodendritic polyamines from OEI and various cores (C6, C4, C2) in DMSO at 22°C/24h. The resulting structures are defined by the core and the number of OEI units (n).

E: X = ethanolamine
 $NH(CH_2)_2OH$

Sp: X = spermidine
 $NH(CH_2)_3NH(CH_2)_4NH_2$

S: X = spermine
 $NH(CH_2)_3NH(CH_2)_4NH(CH_2)_3NH_2$

O: X = OEI
 $NH(CH_2)_3NH_2$

C 6 → HD-core
 C 4 → BD-core
 C 2 → ED-core

Verena Russ et al, *Gene Therapy* 2008

nsm Biocompatibility Studies of HD O *in vivo* **LMU**

A/J mice, polyplexes in HBG buffer
i.v. application, 50 µg pCMVLuc per mouse

- no necrotic changes in liver tissue
- liver enzyme levels similar to control

	(AST + ALT)/GLDH	AP
buffer	5,3	171
HD O c/p 1	6,3	189
LPEI c/p 0.8	11,7	190
BPEI c/p 0.8	17,3	253

- HDO pseudodendrimer mediates *in vivo* gene transfer to tumor tissue with high biocompatibility

➤ Encouraging starting point for degradable polymers with defined G2 and G3 dendrimer cores

Verena Russ et al, *Gene Therapy* 2008

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Tumor targeted siRNA therapy

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nsm OEI-HD Polyplexes for siRNA Delivery **LMU**

Legend: □ Luc siRNA, ■ siCONTROL

Luciferase expression (% of control)

PEI 22: 0.4/1, 0.8/1, 1.2/1, 2/1
PEI 25: 0.4/1, 0.8/1, 1.2/1, 2/1
OEI-HD: 1/1, 2/1, 4/1

Nicole Tietze, Jaro Pelisek et al *Oligonucleotides* 2008
Peter Tarcha et al *Biomaterials* 2007
Julia Kloeckner et al *Bioconjug. Chem.* 2006

nsm Transferrin-coated OEI-HD Polyplexes for siRNA Delivery into Tumors **LMU**

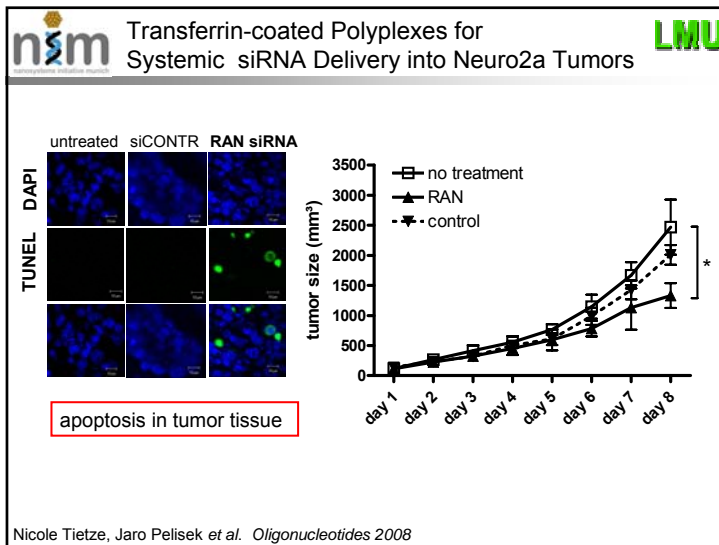
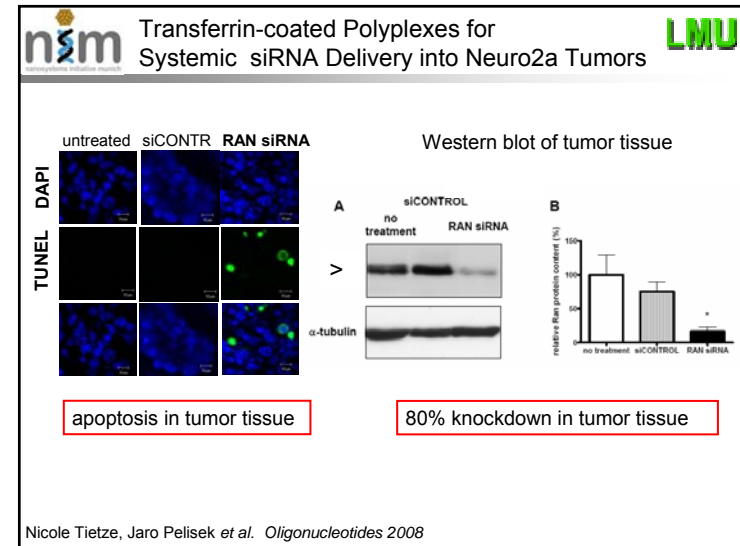
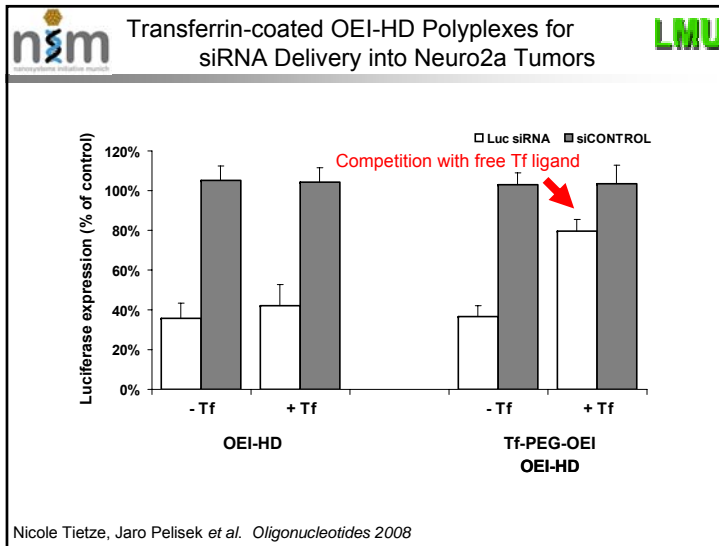
Legend: □ Tf-PEG-OEI, ■ PEG-OEI

% of Cys5 positive cells

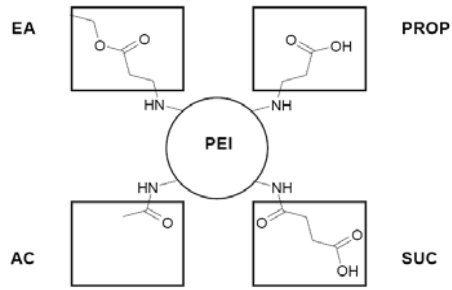
Time [h]: 1, 4, 8, 12, 24

Faster internalization by ligand Tf

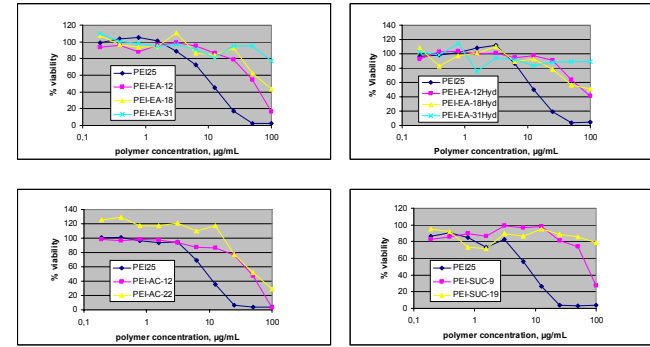
Nicole Tietze, Jaro Pelisek et al. *Oligonucleotides* 2008



- Polyethylenimine for siRNA delivery ?
- Moderate efficiency of PEI (Grayson/Putnam 2006, others)
 - Depending on type (linear/branched) and size of PEI, e.g. LMW purified branched PEI (Aigner 2006)
 - *In vitro* vs. *in vivo* efficiency
 - Stability issue (→ sticky siRNA, Bolcato Bellemin/Behr 2007)
 - Intracellular release issue (LMW PEI for mRNA delivery, Bettinger 2001)

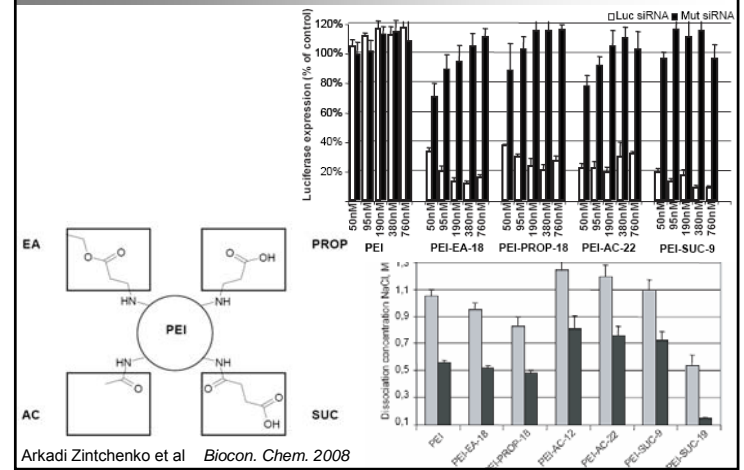
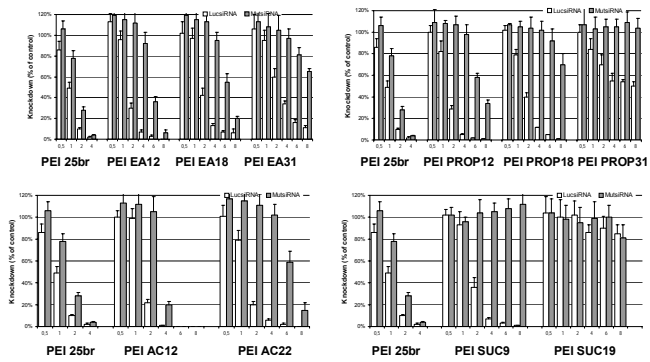


Arkadi Zintchenko et al. *Biocon. Chem.* 2008



Arkadi Zintchenko et al. *Biocon. Chem.* 2008

Neuro2A cells



Arkadi Zintchenko et al. *Biocon. Chem.* 2008

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Improved Polyplexes:

- Better Defined Polymers
- **Better Endosomal Escape**
- Extracellular stabilization
Intracellular disassembly

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nsm Programmed Nanosystems – ‘Synthetic Viruses’ **LMU**

- Pre-programming to alter their properties during drug delivery

Chemical Molecular Sensor Responding to Biological Trigger
Physical Physical

Shielding function (blood circulation)
Targeting function
Triggered release (at target site/cell)

nsm Photochemical Internalization (PCI): Improved Endosomal Release **LMU**

- Sensor = Photosensitizer Trigger = Light

EGF polyplex Sensor = amphiphilic photosensitizer (PS) Stimulus = Light

endocytosis LIGHT $PS \xrightarrow{h\nu} {}^1PS^*$ ${}^1PS^* + O_2 \rightarrow {}^1O_2$

Cell Type	- PCI	+ PCI
PEI22	~10 ⁵	~10 ⁶
EGF/OE1	~10 ⁵	~10 ⁶

Wagner + Kristian Berg lab (Oslo)
J. Kloeckner et al *J Drug Targ* 2004; *JCR* 2006

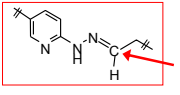
nsm Photochemical Internalization (PCI): Improved Endosomal Release **LMU**


- Sensor = Photosensitizer Trigger = Light

Nadia Ruthardt, Karla de Bruin et al *JCR* 2008
CeNS collaboration Christoph Bräuchle lab

Dynamic Polymers : PEG Deshielding / Endosomal Escape

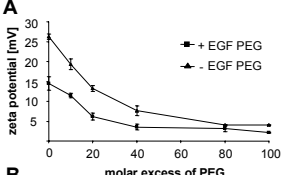
• Sensors = pH-labile bonds Trigger = endosomal pH

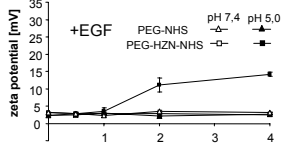
Sensor 1 =  **Stimulus = endosomal pH 5.5**

Sensor 2 =  **Melittin**

pLL, PEI polyplexes
Walker *Mol. Ther.* 2005
Fella *EJPS* 2008
Meyer *JGM* 2007

pH-reversible bifunctional PEG : HZN Linkage

A 

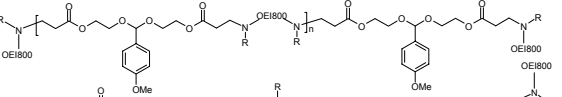
B 

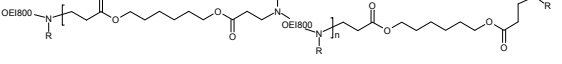
Diffusion times by fluorescence correlation spectroscopy (FCS)

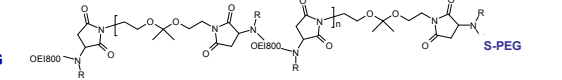
	pH	Diffusion time (µs)
Cysteamine-modified Alexa-NHS	7.4	31 +/- 5
Alexa-SS-PEG-NHS	7.4	193 +/- 18
Alexa-SS-PEG-HZN-NHS	7.4	196 +/- 21
DNA/BPEI, Alexa-SS-PEG-NHS	7.4	1826 +/- 560
DNA/BPEI, Alexa-SS-PEG-NHS	5.0	1988 +/- 1209
DNA/BPEI, Alexa-SS-PEG-HZN-NHS	7.4	1713 +/- 314
DNA/BPEI, Alexa-SS-PEG-HZN-NHS	5.0	192 +/- 19

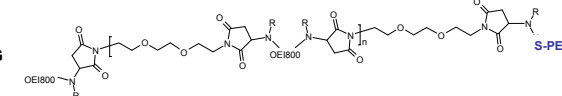
C. Fella et al *EJPS* 2008

Acid-labile Acetal vs Acid-stable Polymers

OEI-BAA 

LT-OEI-HD 

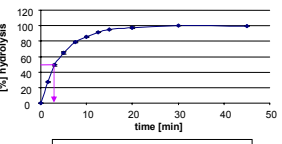
OEI-MK-PEG  **S-PEG**

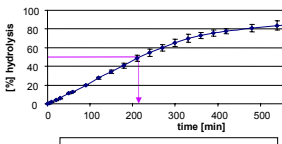
OEI-BM-PEG  **S-PEG**

***OEI *OEI *OEI** $\xrightarrow{\text{pH 5.5}}$ **OEI OEI OEI**

Veronika Knorr et al 2008

Degradable Acetal Polymers

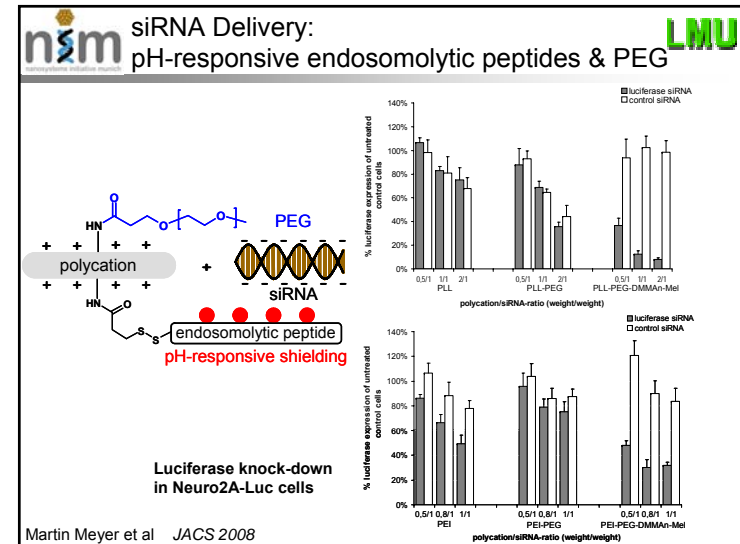
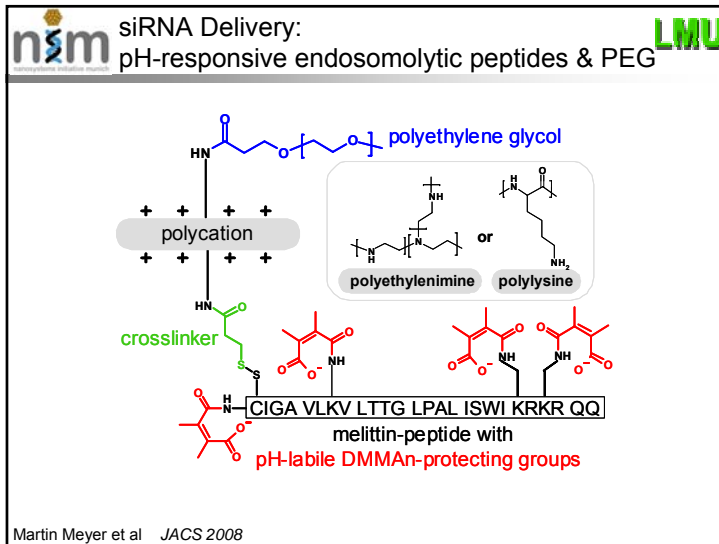
pH 5.0 hydrolysis 

pH 7.4 hydrolysis 

OEI-BAA:
pH 7.4 ~ 3 ½ h (at 37°C)
pH 5.0 ~3 min

OEI-MK:
pH 7.4 ~ 5 h (at 37°C)
pH 5.0 degraded after 15 min

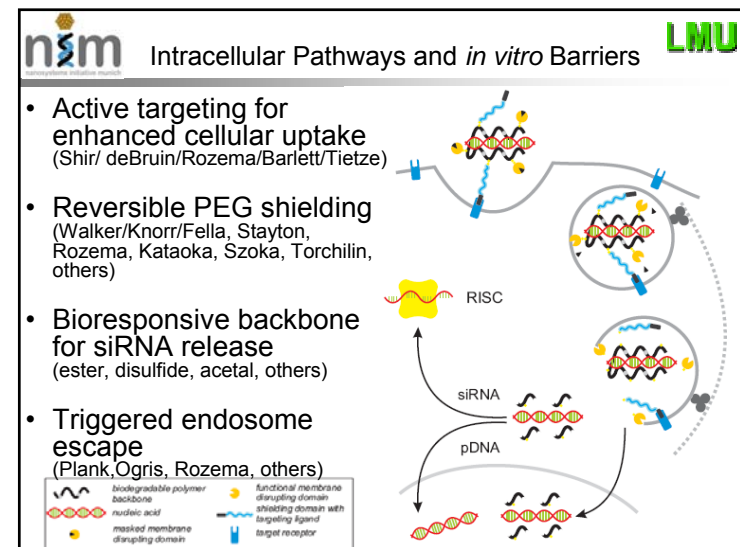
Veronika Knorr et al *Bioconjug. Chem.* 2008



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Conclusions and Prospects

Schaffert & Wagner, *Gene Therapy* 2008
Gene therapy progress and prospects: synthetic polymer-based systems



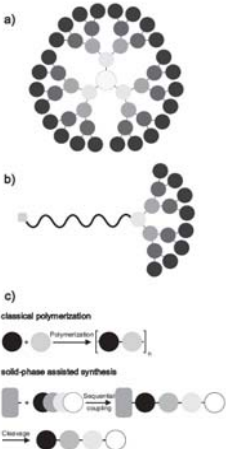
nm Extracellular Pathways and *in vivo* Barriers **LMU**

- Avoid unspecific interactions
 - shielding (PEG, various molecules)
- Avoid extracellular polyplex dissociation
 - (e.g. Burke/Pun 2008 ; Buyens JCR 2008)
 - sticky siRNA (Bolcato-Bellemin/Behr 2007)
 - bioreversible lateral stabilization (Oupicky 2001, Neu 2007)
 - bioreversible covalent conjugates (Rozema/Wolff 2007)
- Avoid siRNA degradation
 - chemically stabilized siRNA
 - protective polymers
- Reduce toxicity
 - biodegradable, balanced systems



nm Polymers for siRNA – Chemistry & Development **LMU**

- Dynamic, bioresponsive 'programmed' polymers which adapt to delivery barriers
 - extracellular stabilization
 - intracellular disassembly
- Improved chemistry will provide
 - monodisperse polymers with uniform size and topology (solid-phase, dendrons, dendrimers)
 - multifunctional sequence defined polymers (e.g. Hartmann/Börner 2008)



Manfred Ogris, group leader vectorology

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